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Synthesis of 4b,5,10a,11-tetrahydroindeno[1,2-*b*]quinolin-10-ones from Baylis–Hillman adducts

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Abstract—We developed an efficient synthetic method for indenoquinoline skeletons from Baylis–Hillman adducts. 4b,5,10a,11-Tetrahydroindeno[1,2-*b*]quinolin-10-ones and 7*H*-indeno[2,1-*c*]quinolines were prepared from Baylis–Hillman adducts in polyphosphoric acid.

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Recently, Basavaiah et al. have reported the synthesis of 3-benzylidenechroman-4-ones from Baylis–Hillman adducts via the Friedel–Crafts acylation reaction as the key step.¹ During the continuing efforts for the construction of various types of heterocyclic compounds from Baylis–Hillman adducts,² we envisioned that we could synthesize the 7*H*-indeno[2,1-*c*]quinoline ring system from the Baylis–Hillman adducts via successive Friedel–Crafts type reaction (Scheme 1). As shown, Friedel–Crafts acylation of cinnamic acid derivative to form 3-benzylidene-2,3-dihydro-4(1*H*)quinolone and the following dehydrative cyclization would furnish the desired 7*H*-indeno[2,1-*c*]quinoline.

Indenoquinoline derivatives^{3–6} showed a wide range of biological activities such as 5-HT-receptor binding activity,^{3b} anti-inflammatory activity,^{4c} and also act as anti-tumor agents,^{3c,5b,d} inhibitor for steroid reductase,^{3d} acetylcholonesterase inhibitors,^{5e} and antimalarials.^{5a} These compounds are frequently synthesized via the aza-Bergman cyclization^{5c} or via the imino Diels–Alder reaction.^{4a,b}

The required starting materials 3a-e were prepared in moderate to good yields as shown in Scheme 2 by simply heating the corresponding anilines 2 and the acetates of the Baylis-Hillman adducts 1 in THF for 14–18h. *E*form of 3 was generated as the major product as reported in a similar system^{2,7} and we could isolate the *E*-form in pure state. The carboxylic acid derivatives **4a–e** were made by KOH-assisted hydrolysis of **3a–e** (Scheme 2).⁷

With **3** and **4** in our hands we examined the possibility for the synthesis of 7*H*-indeno[2,1-*c*]quinoline derivatives. Initially, we examined the reaction of **3a** in various reaction conditions. As examples, we examined the following conditions including heating in TFA, heating in CH₂Cl₂ in the presence of TFAA,¹ heating in benzene with H₂SO₄, and heating in PPA. Among the examined reaction conditions we found some major components on TLC when we conducted the reaction in polyphosphoric acid (PPA) at around 120 °C.⁸ We isolated three compounds, and after careful examination of the spectroscopic data, we finally found that the compounds have the structures of **6a** (3%), **7a** (62%), and **8a** (4%) (Scheme 3).⁹

The structure of **7a** was confirmed by ¹H, ¹³C, IR, mass, and NOE experiments (Fig. 1).⁹ Irradiation of the proton H_a (δ = 4.65 ppm) showed increments of the intensities of H_b and H_e in 1.9% and 1.1%, respectively. Irradiation of the H_b proton (δ = 3.21 ppm) showed enhancements of the intensities of H_a and H_c in 2.4% and 1.6%, respectively. From the NOE experiments, we could conclude that the stereochemistry of ring junction of **7a** as *cis*. In the ¹H NMR spectrum of **8a**, singlet of H-6 appeared at 9.14 ppm. One doublet and two doublets of doublet (presumably corresponding to H-1, H-11, H-4) appeared at 8.43, 8.24, and 8.69 ppm, respectively. Other peaks were overlapped and appeared at around 7.46–7.78 ppm. The postulated reaction mechanism

Keywords: Tetrahydroindeno[1,2-*b*]quinolin-10-ones; 7*H*-Indeno[2,1-*c*]quinolines; Baylis–Hillman adducts; Polyphosphoric acid.

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Scheme 1. Synthetic rationale for 7H-indeno[2,1-c]quinoline.



Scheme 2. Synthesis of starting materials 3 and 4.



Scheme 3. Synthesis of indenoquinolines 7a and 8a.

for the formation of 6a and 7a is depicted in Scheme 3. The double bond of 3a can be isomerized to form 5a as already reported in similar systems by us and other groups.¹⁰ In our acidic medium, 5a can be protonated

to generate the corresponding iminium ion 5a'. The intramolecular aromatic Mannich type reaction¹¹ of 5a' furnished 6a, which can be converted to the 4b,5,10a,11-tetrahydroindeno[1,2-*b*]quinolin-10-one



Figure 1. NOE and coupling constant data of 7a.

(7a) via the following Friedel–Crafts acylation. The formation and isolation of 6a provided us with definite proof for the reaction mechanism of 7a. In a suitably controlled reaction conditions (PPA, 90 °C, 8h) we ob-

Table 1. Synthesis of 7a-f and 8a-e



Scheme 4.

tained the intermediate **6a** as the major product (73%) together with 9% of **7a**. From the reaction of **6a** at elevated temperature (120 °C, 16h) the formation of **7a** from **6a** could be confirmed as in Scheme 4.

As a next trial, in order to find better conditions for the formation of **8a**, we examined a variety of conditions. As shown in Scheme 1, initially we expected that Friedel–Crafts acylation of **4a** might proceed without difficulty to produce 3-benzylidene-2,3-dihydro-4(1*H*) quinolone.¹ If the double bond of 3-benzylidene-2,3-dihydro-4(1*H*)quinolone (**4a**) could be isomerized into its

Entry	$3 \text{ or } 4^{a}$	Conditions	Products		Yield (%), mp (°C)
1 2	3a 4a	PPA 120°C, 16h PPA 170°C, 1h	O HN 7a		7a (62), (149–150), 8a (4) ^b 7a , [°] 8a (18)
3 4	3b 4b	PPA 120°C, 15h PPA 170°C, 30min		CH ₃ 8b	7b (55), (144–145), 8b ^{b,c} 7b (4), 8b (17)
5 6	3c 4c	PPA 120°C, 13h PPA 170°C, 1h			7c (50), (182–183), 8c ^{b,c} 7c (3), 8c (14)
7 8	3d 4d	PPA 120°C, 18h PPA 170°C, 30min	H ₃ C HN HN 7d	CH ₃ Sd N	7d (54), (156–157), 8d ^{b,c} 7d (4), 8d (22)
9 10	3e 4e	PPA 120°C, 17h PPA 170°C, 1h		CI 8e	7e (52), (103–104), 8e ^{b,c} 7e, ^c 8e (13)
11	3b -Z ^d ♀	PPA 120°C, 15h	7b	0	7b (57), (144–145), 8b ^c
12	3f°	PPA 120°C, 15h	H ₃ C H ₃ C		7f (53), (150–151)

^a E-isomer.

^b Oil.

^c Trace amounts on TLC observation.

^d Z-isomer.

^e E/Z = 4:1 mixture.

endo-position via 1,3-hydrogen shift under the acidic conditions,¹⁰ then, the following Friedel–Crafts reaction and dehydration would afford the desired 7*H*-indeno-[2,1-*c*]quinoline (**8a**). But, unfortunately, the reaction of **4a** in PPA showed the formation of complex mixtures as compared in the reaction of **3a** in PPA (vide supra). When we used **4a** as the starting material we could obtain low yield of **7a** (<5%) and **8a** (18%) by using PPA at around 170 °C. Until now, this is the best result for the synthesis of **8a**. Some representative results for the synthesis of **7** and **8** are summarized in Table 1.

We used para-substituted arenes both at the Baylis-Hillman moiety and at the aniline moiety in order to propure regioisomeric products. duce only The stereochemistry of the double bond of starting materials are E-form in all cases except entry 11.12 We separated the Z-form of **3b** and examined the reaction in PPA (entry 11). The reaction of 3b-Z in PPA gave the same product 7b in 57% yield as in the reaction of 3b-E as expected. This means that the configuration of the double bond of 3 did not affect the whole reaction due to the involvement of the double bond isomerization step during the reaction. In another experiment, the reaction of **3f** (E/Z = 4:1 in this case, made from *N*-methylaniline) afforded the corresponding 7f in a similar yield (53%, entry 12).

In summary, we have disclosed an efficient entry for the synthesis of valuable indenoquinoline skeletons from the easily available Baylis–Hillman adducts in PPA medium. We are currently searching the improved conditions for the synthesis of 7H-indeno[2,1-c]quinolines.

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- 9. Typical procedure for the synthesis of 6a, 7a, and 8a: The reaction of 1a and aniline (3 equiv) in refluxing THF (15h) gave 3a in 69% yield. A stirred solution of 3a (534mg, 2mmol) in PPA (6mL) was heated to 120 °C for 16h. After pouring the reaction mixture into cold water the aqueous phase was neutralized cautiously with NaHCO₃. Extraction with ether, removal of solvent, and flash column chromatographic separation process (hexanes/ether, 2:1) gave 6a (16mg, 3%), 7a (292mg, 62%), and 8a (18mg, 4%). Compound 6a: ¹H NMR (CDCl₃) δ 2.97–3.04 (m, 1H), 3.57 (s, 3H), 3.32-3.60 (m, 2H), 3.95 (br s, 1H), 4.48 (d, J = 7.2 Hz, 1 H), 6.55–6.62 (m, 2H), 6.72 (d, J = 7.5 Hz, 1H), 7.00 (t, J = 7.8 Hz, 1H), 7.11–7.31 (m, 5H); ¹³C NMR $(CDCl_3)$ δ 41.90, 45.26, 46.83, 51.83, 114.40, 117.76, 122.52, 126.61, 127.30, 128.38, 128.93, 130.47, 143.86, 144.42, 173.45. Compound 7a: IR (neat) 3340, 1709 cm⁻¹; ¹H NMR (CDCl₃) δ 3.21 (ddd, J = 8.1, 4.8, and 2.7 Hz, 1H), 3.31 (dd, J = 11.1 and 4.8 Hz, 1H), 3.65 (br s, 1H), 3.74 (dd, J = 11.1 and 2.7 Hz, 1H), 4.65 (d, J = 8.1 Hz, 1H), 6.52 (dd, J = 7.8 and 1.2 Hz, 1H), 6.85 (td, J = 7.5 and 1.2 Hz, 1H), 7.01 (td, J = 7.8 and 1.2 Hz, 1H), 7.25–7.36 (m, 1H), 7.48–7.63 (m, 3H), 7.73 (d, J = 7.5 Hz, 1H); ¹³C NMR (CDCl₃) & 40.23, 44.35, 50.28, 115.90, 119.60, 123.28, 124.43, 126.25, 127.24, 127.74, 129.20, 135.26, 135.65, 146.54, 156.52, 207.77; EI-MS (70eV) m/z (rel. intensity) 77 (12), 130 (31), 178 (14), 206 (24), 218 (30), 234 (67), 235

 $(M^+, 100).$

Compound **8a**: ¹H NMR (CDCl₃) δ 4.08 (s, 2H), 7.46–7.58 (m, 2H), 7.65–7.78 (m, 3H), 8.24 (dd, J = 8.4 and 1.2 Hz, 1H), 8.43 (d, J = 7.5 Hz, 1H), 8.69 (dd, J = 8.4 and 1.2 Hz, 1H), 9.14 (s, 1H); ¹³C NMR (CDCl₃) δ 35.72, 123.65, 124.43, 124.64, 125.47, 126.83, 127.38, 128.22, 128.43, 130.51, 135.85, 140.70, 144.47, 144.97, 147.63, 147.91; MS (MALDI-TOF) 217.4 (M⁺).

The other experiments were carried out analogously and the spectroscopic data of the prepared compounds are as follows.

Compound **7b**: IR (neat) 3344, 1712 cm^{-1} ; ¹H NMR (CDCl₃) δ 2.32 (s, 3H), 3.17–3.22 (m, 1H), 3.26–3.31 (m, 1H), 3.52 (br s, 1H), 3.74 (dd, J = 11.1 and 2.7Hz, 1H), 4.60 (d, J = 7.8 Hz, 1H), 6.44 (d, J = 7.8 Hz, 1H), 6.83 (dd, J = 7.8 and 1.5Hz, 1H), 7.31–7.36 (m, 2H), 7.53–7.63 (m, 2H), 7.73 (d, J = 7.5 Hz, 1H); ¹³C NMR (CDCl₃) δ 20.71, 40.29, 44.68, 50.26, 115.83, 123.24, 124.45, 126.26, 127.70, 127.86, 128.88, 129.60, 135.26, 135.70, 144.12, 156.55, 208.02; EI-MS (70 eV) m/z (rel. intensity) 144 (27), 234 (24), 248 (64), 249 (M⁺, 100).

Compound 7c: IR (neat) 3340, 1712 cm^{-1} ; ¹H NMR (CDCl₃) δ 3.18–3.31 (m, 1H), 3.26–3.32 (m, 1H), 3.68 (br s, 1H), 3.76 (dd, J = 11.1 and 2.7Hz, 1H), 4.58 (d, J = 8.1 Hz, 1H), 6.46 (d, J = 8.4 Hz, 1H), 6.96 (dd, J = 8.1 and 2.1 Hz, 1H), 7.34–7.40 (m, 1H), 7.46 (dd, J = 2.4 and 0.6 Hz, 1H), 7.59–7.61 (m, 2H), 7.74 (d, J = 7.5 Hz, 1H); ¹³C NMR (CDCl₃) δ 40.09, 44.19, 49.96, 117.01, 123.42, 123.96, 125.94, 126.19, 127.15, 128.04, 128.80, 135.47, 135.64, 145.13, 155.69, 207.18; EI-MS (70 eV) *m/z* (rel. intensity) 164 (32), 204 (26), 233 (24), 251 (21), 269 (M⁺, 100).

Compound 7d: IR (neat) 3363, 1709 cm^{-1} ; ¹H NMR (CDCl₃) δ 2.36 (s, 3H), 3.19–3.24 (m, 1H), 3.28–3.34 (m, 1H), 3.62 (br s, 1H), 3.74 (dd, J = 10.8 and 2.7Hz, 1H), 4.61 (d, J = 8.1 Hz, 1H), 6.52 (dd, J = 7.8 and 1.2Hz, 1H), 6.84 (td, J = 7.2 and 1.2Hz, 1H), 7.01 (td, J = 7.8 and 1.5Hz, 1H), 7.37 (dd, J = 8.1 and 1.2Hz, 1H), 7.47–7.53 (m, 3H); ¹³C NMR (CDCl₃) δ 20.98, 39.92, 44.47, 50.64, 115.89, 119.60, 123.18, 124.75, 125.93, 127.16, 129.17, 135.82, 136.56, 137.78, 146.49, 154.02, 207.87; EI-MS (70 eV) m/z (rel. intensity) 130 (31), 204 (17), 220 (17), 232 (39), 249 (M⁺, 100).

Compound 7e: IR (neat) 3351, 1709 cm^{-1} ; ¹H NMR (CDCl₃) δ 3.20–3.31 (m, 2H), 3.63 (br s, 1H), 3.73 (dd, J = 10.8 and 2.4Hz, 1H), 4.60 (d, J = 7.8 Hz, 1H), 6.52 (dd, J = 7.8 and 1.2Hz, 1H), 6.85 (td, J = 7.8 and 1.2Hz, 1H), 7.02 (td, J = 7.8 and 1.2Hz, 1H), 7.43–7.53 (m, 3H), 7.66 (d, J = 2.1 Hz, 1H); ¹³C NMR (CDCl₃) δ 39.88, 44.44, 50.74, 115.98, 119.75, 122.89, 123.89, 127.42, 127.54, 129.00, 134.15, 135.18, 137.02, 146.51, 154.56, 206.58; EI-MS (70eV) m/z (rel. intensity) 130 (33), 204 (30), 233 (20), 252 (36), 269 (M⁺, 100).

Compound **7f**: IR (neat) 1712 cm^{-1} ; ¹H NMR (CDCl₃) δ 2.61 (s, 3H), 3.02 (dd, J = 11.1 and 5.1 Hz, 1H), 3.09–3.15 (m, 1H), 3.51 (dd, J = 11.1 and 2.1 Hz, 1H), 4.55 (d, J = 8.1 Hz, 1H), 6.53 (dd, J = 8.1 and 0.9 Hz, 1H), 6.81 (td, J = 7.5 and 1.2 Hz, 1H), 7.05 (td, J = 7.8 and 1.5 Hz, 1H), 7.21–7.27 (m, 1H), 7.40–7.50 (m, 3H), 7.64 (d, J = 7.8 Hz,

112.78, 119.35, 123.35, 126.34, 126.42, 127.78, 127.90, 128.98, 135.44, 136.02, 149.16, 156.78, 208.37; EI-MS (70 eV) m/z (rel. intensity) 144 (29), 178 (11), 204 (14), 220 (17), 232 (29), 249 (M⁺, 100). Compound 8b: ¹H NMR (CDCl₃) & 2.66 (s, 3H), 4.06 (s, 2H), 7.45–7.59 (m, 3H), 7.70 (d, J = 6.0 Hz, 1H), 8.12 (d, J = 6.0 Hz, 1H), 8.42–8.44 (m, 2H), 9.06 (s, 1H); ¹³C NMR $(CDCl_3)$ δ 22.34, 35.92, 122.90, 124.64, 124.89, 125.66, 127.52, 128.28, 130.38, 130.87, 136.11, 136.98, 141.09, 143.98, 145.17, 146.71, 146.95; EI-MS (70 eV) *m/z* (rel. intensity) 115 (5), 189 (5), 202 (11), 216 (17), 231 (M⁺, 100)Compound 8c: ¹H NMR (CDCl₃) δ 4.09 (s, 2H), 7.48–7.61 (m, 2H), 7.67–7.74 (m, 2H), 8.17 (d, J = 9.0 Hz, 1H), 8.37 (d, J = 7.2 Hz, 1H), 8.65 (d, J = 2.4 Hz, 1H), 9.12 (s, 1H); ¹³C NMR (CDCl₃) δ 35.73, 122.80, 124.19, 125.10, 125.56, 127.56, 128.55, 129.36, 132.02, 132.74, 136.69, 140.09, 143.80, 144.85, 146.30, 147.79; EI-MS (70eV) m/z (rel. intensity) 94 (10), 125 (9), 189 (20), 216 (96), 251 (M⁺, 100).Compound 8d: ¹H NMR (CDCl₃) δ 2.57 (s, 3H), 4.03 (s, 2H), 7.31 (dd, J = 7.5 and 0.6Hz, 1H), 7.59 (d, J = 7.5 Hz, 1H), 7.66-7.78 (m, 2H), 8.22-8.25 (m, 2H), 8.71 (dd, J = 8.4 and 1.2 Hz, 1H), 9.11 (s, 1H); ¹³C NMR (CDCl₃) δ 21.84, 35.35, 123.75, 124.71, 125.06, 125.10, 126.74, 128.39, 129.24, 130.50, 136.28, 137.05, 140.94, 142.13, 144.53, 147.67, 147.92; EI-MS (70eV) m/z (rel. intensity) 101 (5), 189 (6), 202 (15), 216 (23), 231 (M⁺, 100).

1H); ¹³C NMR (CDCl₃) δ 39.74, 41.09, 50.66, 53.99,

Compound **8e**: IR (neat) 2924, 1562, 1508 cm⁻¹; ¹H NMR (CDCl₃) δ 4.05 (s, 2H), 7.43–7.80 (m, 4H), 8.24 (d, J = 8.1 Hz, 1H), 8.37 (s, 1H), 8.58 (d, J = 8.1 Hz, 1H), 9.12 (s, 1H); ¹³C NMR (CDCl₃) δ 35.60, 123.52, 124.57, 124.73, 126.54, 127.46, 128.40, 128.93, 130.84, 133.58, 136.68, 142.52, 143.27, 143.54, 147.78, 148.11.

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- 12. As already published,^{2,7} the $S_N 2'$ reaction of the Baylis– Hillman acetate and nucleophiles including aniline afforded the *E*-form product as the major component. The corresponding *Z*-form was generated in about 5–20% depending on the substrates. Thus, we scaled up the reaction and obtained the minor component **3b**-*Z*.